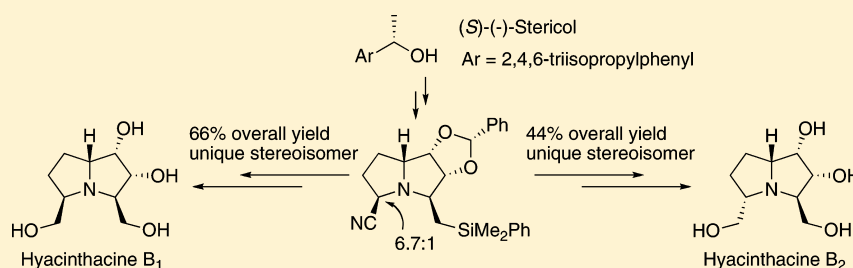


Asymmetric Approach to Hyacinthacines B₁ and B₂

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S Supporting Information



ABSTRACT: Naturally occurring hyacinthacines B₁ and B₂ have been prepared from a common, easily available, advanced intermediate. The approach features several highly stereoselective transformations: inter alia, a dichloroketene–enol ether [2 + 2] cycloaddition, a Bruylants alkylation, and an amino–nitrile alkylation–reduction.

INTRODUCTION

Glycosidase inhibition has received considerable attention over the past several years, mainly because glycosidases, which catalyze the hydrolysis of oligosaccharides and glycoconjugates, are involved in an array of diverse biological processes.¹ Inhibitors of these enzymes can thus be viewed as potential drugs against various human diseases.² With relatively few compounds presently in clinical use, however, research aimed at finding new inhibitors continues unabated.³

The iminosugars⁴ are currently among the most studied glycosidase inhibitors, and the hyacinthacines, which are polyhydroxylated pyrrolizidines, figure prominently within this group (Figure 1).⁵ Since the isolation of the first of the hyacinthacine alkaloids by Asano and co-workers in 1999,⁶ more than 20 have been identified. These iminosugars are characterized by the presence of an hydroxymethyl substituent at C-3 and are divided into 3 groups, A, B, and C, on the basis

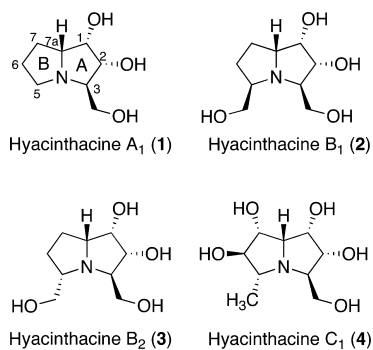


Figure 1. Examples of hyacinthacines.

of the number of hydroxyl and hydroxymethyl substituents on ring B of the pyrrolizidine system.⁷

The hyacinthacines have been shown to possess strong, selective biological activities, which, coupled with their compact, highly functionalized structures, make them worthwhile synthetic targets. Not surprisingly, synthetic efforts toward the hyacinthacines have so far focused primarily on the preparation of the simpler group A members, in racemic form as well as natural form (using chiral pool or enzymatic techniques).^{8,9} A few years ago, we reported the first nonchiral pool/nonenzymatic approach to natural hyacinthacine A₁,¹⁰ which relied on a strategy based on asymmetric dichloroketene–chiral enol ether cycloaddition,¹¹ previously applied^{12–14} for the preparation of a variety of other alkaloids.¹⁵ We now describe in detail the extension of this work to access the more structurally complex hyacinthacines B₁ and B₂.¹⁶ These alkaloids, isolated in low yield (22 mg/kg and 5 mg/kg, respectively) by Asano and co-workers from bulbs of *Scilla campanulata*, have been shown to be selective inhibitors of β -glucosidase and β -galactosidase.⁶ Their structures are characterized by the presence of five stereocenters and two synthetically challenging hydroxymethyls at C-3 and C-5. To the best of our knowledge, only one other approach to these alkaloids, which employed L-pyroglutamic acid for chirality, has been reported.^{9a}

RESULTS AND DISCUSSION

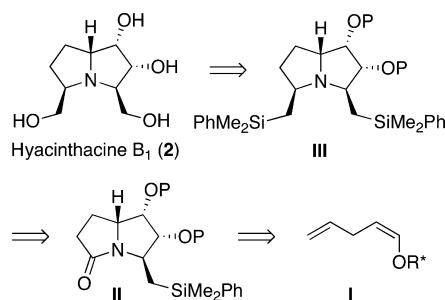
Synthesis of Hyacinthacine B₁. It was envisaged that hyacinthacine B₁ would be obtained from the disilyl derivative

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III through an uncommon double Tamao–Fleming oxidation to generate in a single step the requisite hydroxymethyl groups (Scheme 1). The C-5 dimethylphenylsilylmethyl substituent

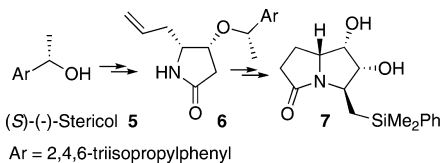
Scheme 1. Retrosynthesis of (+)-Hyacinthacine B₁



might be introduced stereoselectively through a Bruylants reaction¹⁷ of the amino-nitrile obtained through reductive cyanation of pyrrolizidinone II. Significantly, pyrrolizidinone II seemed to offer as well several means to access hyacinthacine B₂. The unprotected pyrrolizidinone II (P = H) had earlier been prepared from enol ether I (R*OH = Stericol).¹⁰

The synthesis started from the commercially available chiral auxiliary (S)-(-)-Stericol 5,¹⁸ which was transformed in a highly stereoselective manner into pyrrolizidinone 7 via lactam 6^{14,19} (Scheme 2).¹⁰ The key diastereoselective steps in this preparation included a [2 + 2] cycloaddition of dichloroketene to the Stericol-derived dienol ether, a Bruylants-like addition of a silylmethyl substituent, and an *endo*-selective dihydroxylation.

Scheme 2. Preparation¹⁰ of Pyrrolizidinone 7

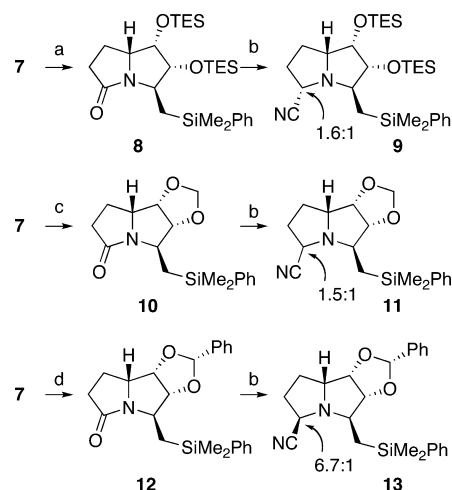


The protection of the *cis* hydroxyls in 7 was expected to be crucially important in the approach because of not only introduction–removal considerations, but also the attendant steric hindrance that could play a significant role in the subsequent C-5 alkylation. Thus, different groups were examined. Protection with triethylsilyl groups and a methylidene group proceeded smoothly to give lactams 8 and 10 in 94 and 88% yields, respectively (Scheme 3).

Benzylidene protection of 7 was particularly interesting in that a single, crystalline acetal was formed in excellent yield; structural determination by X-ray diffraction analysis²⁰ revealed that the *phenyl* group of the acetal was *endo* in the tricyclic structure (Figure 2). This *endo* stereoselectivity, while at first surprising, is in fact often encountered with cyclic diols²¹ and is believed to result from kinetic acetal formation, with minimization of allylic strain²² (A^{1,3}) in the cyclization of the intermediate oxonium ion.²³

The transformation of the lactams to the corresponding amino-nitriles²⁴ by reductive cyanation, studied initially with the bis-TES derivative 8, turned out to be more challenging than expected: reductive cyanation with the often used DIBAL-H/KCN system²⁵ led invariably to significant amounts of over-reduced material, while the use of Schwartz's reagent with TMSCN²⁶ led primarily to recovered starting material.

Scheme 3. Diol Protection and Amino-Nitrile Formation^a



^aReagents and conditions: (a) TESCl, imidazole, CH₂Cl₂, 20 °C, 94%. (b) DIBAL-H/*n*-BuLi ate complex, THF, 20 °C, then TMSCN, 0 °C, 89% for 9 from 8, 93% for 11 from 10, 97% for 13 from 7 (2 steps). (c) P₂O₅, CH₂(OMe)₂, CH₂Cl₂, 20 °C, 88%. (d) PhCH(OMe)₂, *p*-TSA, CH₂Cl₂, 20 °C.

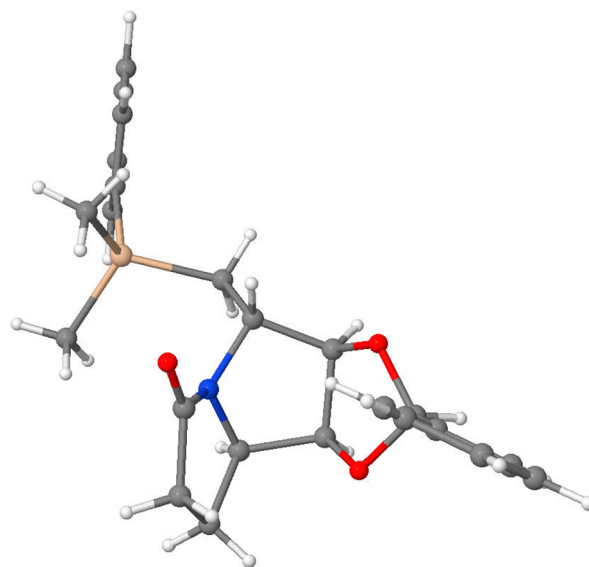
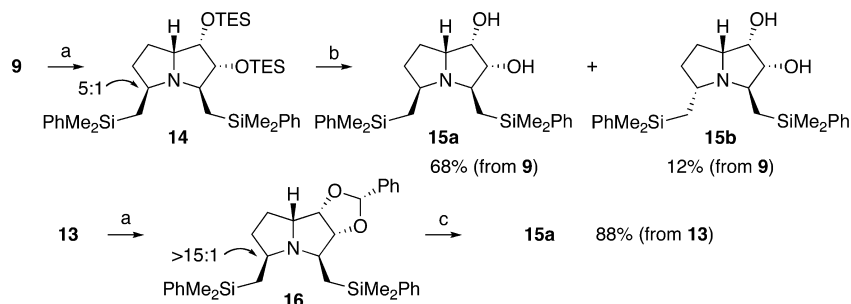


Figure 2. Solid state structure of benzylidene 12.

Fortunately, however, the DIBAL-H/*n*-BuLi ate complex²⁷ smoothly produced the desired hemiaminal, which, following treatment in situ with TMSCN, gave in 89% yield an inconsequential 1.6:1 mixture of the epimeric amino-nitriles 9 (Scheme 3), with the silylmethyl group governing the observed selectivity.²⁸ This procedure applied to the methylidene-protected derivative 10 provided in 89% yield the expected amino-nitrile derivatives 11 as a 1.5:1 mixture (see below for stereochemical assignment), less pronounced than had been expected. When lactam 12 was subjected to the reductive cyanation conditions, however, the amino-nitriles 13 were obtained in 97% overall yield from 7 (2 steps) as a 6.7:1 mixture.²⁹ This facial discrimination proved to be a harbinger of the stereoselectivity of the upcoming alkylation.

With a highly efficient formation of the amino-nitriles in hand, the Bruylants reaction¹⁷ was next examined. The mixture of amino-nitriles 9 reacted with dimethylphenylsilylmethylmag-

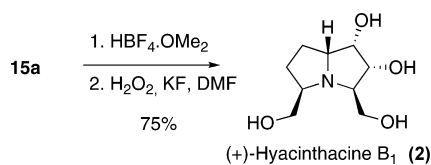
Scheme 4. Bruylants Reaction and Diol Deprotection^a

^aReagents and conditions: (a) $\text{Me}_2\text{PhSiCH}_2\text{MgCl}$, Et_2O –THF, 20 °C. (b) TBAF, THF, 20 °C. (c) TFA (20%), THF– H_2O , 55 °C.

nesium chloride in ether–THF at room temperature to afford a 5:1 epimeric mixture of the corresponding alkylated pyrrolizidines **14** (Scheme 4). A sample of the major isomer, separated with difficulty by silica gel chromatography, provided ^1H NMR data in agreement with the depicted structure. Particularly diagnostic was the strong NOE (10%) between the C-3 and C-5 hydrogens. The TES groups were next removed from **14** by treatment with TBAF and the resultant mixture could now be easily separated by chromatography to give diol **15a** and its C-5 epimer **15b** in 68 and 12% overall yields, respectively, from **9**. For additional proof of the stereochemical assignments from the Bruylants reaction, the pure diol **15b** was silylated to give back the minor C-5 epimer of **14**, which displayed no NOE between the C-3 and C-5 hydrogens, but one between those at C-5 and C-7a.

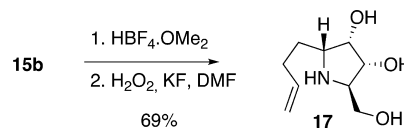
Better yet, when the mixture of amino-nitriles **13** was treated similarly with the same Grignard reagent, a *unique* stereoisomer (**16**) was formed (by ^1H NMR: single signal for the benzylidene acetal hydrogen at 5.70 ppm). Acetal cleavage with aqueous acid then furnished diol **15a** in 88% overall yield from **13** (2 steps).

Silyl oxidation can be performed through several different procedures,³⁰ but few have been successfully used with a potentially oxidizable amino group in the substrate.³¹ With a model substrate, the basic oxidation conditions developed by Smitrovich and Woerpel^{31a} proved insufficiently reactive, whereas treatment with tetrafluoroboric acid, followed by basic workup and hydrogen peroxide oxidation, produced the corresponding protodesilylated amine oxide as the major product. However, by eliminating the basic treatment following the first step so as to keep the nitrogen protonated and thus limit its oxidation, it was found that the desired alcohol was cleanly formed, without significant amounts of the *N*-oxide. To our delight, on application of these conditions, the disilyl derivative **15a** underwent clean double oxidation to afford hyacinthacine *B*₁ in 75% yield, without discernible *N*-oxide formation (Scheme 5). This result is noteworthy since double Tamao–Fleming oxidations are relatively few in the literature³² and, moreover, attendant nitrogen oxidation is often difficult to

Scheme 5. Double Tamao–Fleming Oxidation of **15a** to give (+)-Hyacinthacine *B*₁

suppress.³³ Synthetically derived (+)-hyacinthacine *B*₁ ($[\alpha]_{\text{D}}^{+40}$) was spectroscopically and chromatographically indistinguishable from an authentic sample of the natural product ($[\alpha]_{\text{D}}^{+41.3}$).^{6,34}

Synthesis of Hyacinthacine *B*₂. In order to evaluate the possibility of accessing hyacinthacine *B*₂ through a similar strategy, the minor diol **15b** was subjected to the above oxidation conditions. Surprisingly, this isomer did not undergo transformation into the expected polyhydroxylated pyrrolizidine but suffered an oxidation–fragmentation to generate pyrrolidine **17** in 69% yield after treatment with basic Dowex (Scheme 6).

Scheme 6. Oxidation–Fragmentation of **15b**

This dichotomous behavior of the isomeric disilanes **15a** and **15b** most likely results from conformational biases. Calculations^{35,36} indicate that in **15a** the silane in question enjoys a degree of conformational freedom (a preferred conformation places the silane toward the nitrogen, dihedral angle of -61.4° , Figure 3A), whereas in **15b** it is essentially blocked opposite the nitrogen (dihedral angle of -176.3° , Figure 3B) and favorably disposed to participate in a Grob-type fragmentation.³⁷ A different approach, which did not involve a double Tamao–Fleming oxidation, was obviously required for reaching hyacinthacine *B*₂.

A potential alternative emerged with the observation that the mixture of amino-nitriles **11** (unassigned stereochemistry, Scheme 3), in solution at -30°C , underwent over several days a 1.5:1 to 9:1 modification of the epimeric composition. It was subsequently found that on heating a DMF solution of the epimers, the 9:1 ratio could be attained in only 45 min.³⁸ While these isomers could be separated by silica gel chromatography, their stereo attribution could not be made with absolute confidence. For this reason, $\text{CN} \rightarrow \text{CH}_2\text{OH}$ transformation (methanolysis, reduction), Tamao–Fleming oxidation, and diol deprotection in the major isomer were carried out; unfortunately, hyacinthacine *B*₁ and not *B*₂ was produced. The cyano and silylmethyl group were thus, in fact, *cis* in the major isomer of **11** (Scheme 3), and yet another strategy for the synthesis of hyacinthacine *B*₂ was required.

The complete diastereoselectivity induced above with the benzylidene protecting group suggested that an approach based

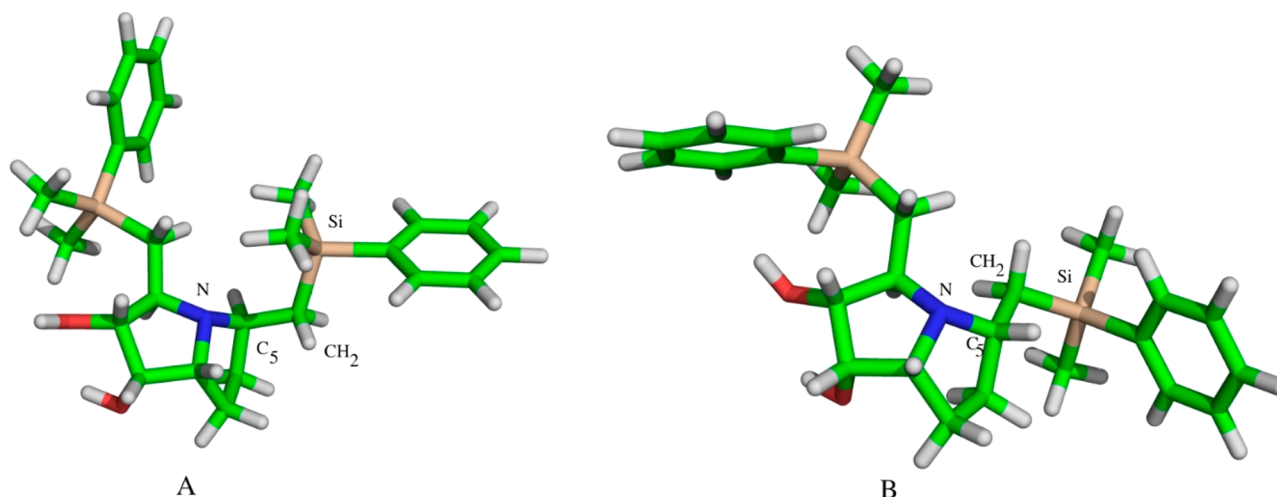
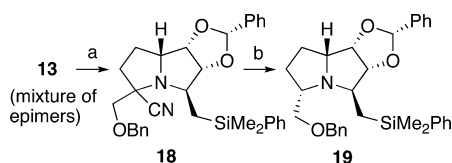


Figure 3. Calculated conformations for 15a and 15b.

on hydride reduction³⁹ of an *alkylated amino-nitrile*⁴⁰ might constitute a viable strategy for preparing hyacinthacine B₂.⁴¹ This plan was particularly attractive in that both hyacinthacines would result from a common, late intermediate. After considerable experimentation, it was found that treatment of amino-nitriles **13** with LDA at low temperature, followed by addition of an excess of benzyl chloromethyl ether, gave the corresponding amino-nitrile **18** in high diastereomeric purity by NMR (Scheme 7). For a successful alkylation, it was necessary

Scheme 7. Alkylation of **13** and Reduction^a

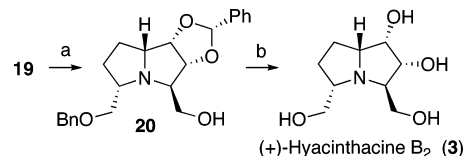


^aReagents and conditions: (a) LDA, THF, $-60\text{ }^{\circ}\text{C}$; benzyl chloromethyl ether, $-60\text{ }^{\circ}\text{C}$. (b) NaBH_4 , MeOH, $0\text{ }^{\circ}\text{C}$, 49% (55% brsm, 2 steps).

to operate with rigorous exclusion of oxygen to avoid reversion of the substrate to lactam **12**.⁴² Although the crude product was not stable to purification, it could be smoothly reduced with sodium borohydride in methanol at $0\text{ }^{\circ}\text{C}$ to afford *uniquely* the desired stereoisomer **19** in 49% yield (55% brsm, 2 steps). Thus, once again, the benzylidene protection allowed total, and in this case opposite, stereocontrol in the introduction of a latent C-5 hydroxymethyl group.

To our temporary dismay, attempted Tamao–Fleming oxidation of silane **19** under the conditions that had previously been used successfully for hyacinthacine B₁ led mainly to degradation. Fortunately, however, the conditions developed by Smitrovich and Woerpel^{31a} were found to be productive and furnished the expected primary alcohol **20** in essentially quantitative yield (Scheme 8). The completion of the synthesis was realized by double deprotection through acidic hydrogenolysis, followed by deprotonation with basic Dowex resin, to provide (+)-hyacinthacine B₂ in 89% yield from silane **19** (2 steps). Synthetically derived (+)-hyacinthacine B₂ provided ¹H and ¹³C NMR data in excellent accord with those described for the naturally derived material.^{6,43}

Scheme 8. Completion of the Synthesis of (+)-Hyacinthacine B₂^a



^aReagents and conditions: (a) KH, *t*-BuOOH, TBAF, DMF, $0\text{ }^{\circ}\text{C}$. (b) H_2 , Pd–C, 6 N HCl, EtOH, $20\text{ }^{\circ}\text{C}$; Dowex (OH[−] form), H₂O, 89% (2 steps).

CONCLUSION

Hyacinthacines B₁ and B₂ have been prepared efficiently from a common, advanced intermediate, which is obtained in enantiopure form through asymmetric cycloaddition methodology. These syntheses are characterized by excellent levels of stereocontrol of the 5 stereogenic centers and high overall yields from the starting chiral auxiliary (6.5 and 4.4%, respectively). Critical to the success of this work is the synthetic versatility of the amino-nitrile moiety, which provides highly selective access to these diastereomeric hyacinthacines. Synthetic studies toward other C-5 substituted hyacinthacines are currently in progress in our laboratory.

EXPERIMENTAL SECTION

General Experimental Details. All experiments were carried out under argon unless otherwise stated. THF and Et₂O were distilled from Na-benzophenone. CH₂Cl₂ and ¹Pr₂NH were distilled over CaH₂ and DMF over CaSO₄. All other materials were directly used as received from commercial sources without purification. NMR spectra were recorded on a Bruker Avance 300, Bruker Avance 400, or Varian U+ 500 spectrometer in chloroform-*d*, unless otherwise stated. All coupling constants have been calculated, when possible, by using the method described by Hoyer et al.⁴⁴ Melting points are uncorrected. IR spectra were recorded as neat samples or in CH₂Cl₂ solution on a Nicolet 397 FT-spectrometer. The mass spectra were recorded on a Nermag R10 mass spectrometer in ESI mode. High-resolution mass spectrometry data were obtained by electrospray ionization with an orbitrap detector.

(5*S*,6*R*,7*S*,7*aR*)-5-((Dimethyl(phenyl)silyl)methyl)-6,7-bis-(triethylsilyloxy)-hexahydro-1*H*-pyrrolizin-3-one (8). To a stirred solution of dihydroxy lactam **7** (0.200 g, 0.655 mmol) and imidazole (0.267 g, 3.92 mmol) in anhydrous dichloromethane (10 mL) at $0\text{ }^{\circ}\text{C}$ was added 0.329 mL (0.295 g, 1.96 mmol) of triethylsilyl

chloride. The mixture was allowed to warm to 20 °C over 30 min and then stirred for 16 h, whereupon water was added. The mixture was extracted with dichloromethane, the combined organic layers were washed with water, dried over anhydrous Na₂SO₄, and filtered, and the filtrate was concentrated to give the crude product. Purification of this material by flash chromatography on silica gel with ethyl acetate in pentane (1:10) afforded 0.330 g (94%) of lactam **8**: [α]_D²⁴ –23 (c 3.2, CHCl₃); IR (film) 2955, 2904, 2873, 1693, 1583, 1458, 1382, 1242, 1181, 1116, 1014 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.34 (s, 3 H), 0.38 (s, 3 H), 0.51–0.63 (m, 12 H), 0.87–0.97 (m, 19 H), 1.17 (dd, *J* = 14.7, 5.2 Hz, 1 H), 1.71–1.80 (m, 1 H), 2.04–2.16 (m, 1 H), 2.17–2.26 (m, 1 H), 2.30–2.41 (m, 1 H), 3.59 (ddd, *J* = 7.0, 5.0, 5.0 Hz, 1 H), 3.75 (dd, *J* = 4.5, 3.5 Hz, 1 H), 3.81–3.87 (m, 1 H), 4.00 (dd, *J* = 5.0, 3.5 Hz, 1 H), 7.32–7.36 (m, 3 H), 7.50–7.55 (m, 2 H); ¹³C NMR (75.5 MHz, CDCl₃) δ –2.8 (CH₃), 2.7 (CH₃), 4.9 (CH₂), 6.7 (CH₃), 6.8 (CH₃), 19.8 (CH₂), 20.3 (CH₂), 32.4 (CH₂), 58.4 (CH), 61.3 (CH), 73.6 (CH), 82.2 (CH), 127.6 (CH), 128.7 (CH), 133.5 (CH), 139.5 (C), 178.4 (C); MS (ESI) *m/z* 556 (MNa⁺); HRMS (ESI) calcd for C₂₈H₅₁N₃O₃Si 556.3069, found 556.3065 (MNa⁺).

(3R,5S,6R,7S,7aR)- and (3S,5S,6R,7S,7aR)-5-((Dimethyl(phenyl)silyl)methyl)-6,7-bis(triethylsilyloxy)-hexahydro-1H-pyrrolizine-3-carbonitriles (9). A stirred solution of lactam **8** (0.122 g, 0.228 mmol) in anhydrous THF (3.0 mL) was treated at 0 °C with a freshly prepared solution of DIBALH-BuLi ate complex (2.5 mL, prepared from 2.8 mL of 1.0 M DIBAL-H in hexanes and 1.1 mL of 2.5 M *n*-BuLi in hexanes in 1.2 mL of anhydrous THF). The reaction mixture was allowed to warm to 20 °C over 30 min and stirred for 16 h, whereupon it was cooled to –60 °C and treated with 0.243 mL (0.193 g, 1.94 mmol) of TMSCN. After being allowed to warm to 0 °C over 1 h, the mixture was diluted with water and then ethyl acetate and filtered through a small pad of Celite. The filtrate was concentrated under reduced pressure to give the crude nitriles, which were purified by flash chromatography with ethyl acetate in pentane (1:10) to give 0.111 g (89%) of the amino-nitriles **9** as a 1.6:1 diastereomeric mixture. A comparable sample was separated for characterization purposes. Major isomer (in mixture with small amount of minor isomer): ¹H NMR (400 MHz, CDCl₃) δ 0.31 (s, 3 H), 0.33 (s, 3 H), 0.60–0.69 (m, 12 H), 0.93–1.03 (m, 18 H), 1.08 (dd, *J* = 15.5, 5.0 Hz, 1 H), 1.25 (dd, *J* = 15.2, 5.0 Hz, 1 H), 1.57–1.65 (m, 1 H), 1.80–1.89 (m, 1 H), 1.99–2.12 (m, 2 H), 3.10–3.14 (m, 1 H), 3.27–3.32 (m, 1 H), 3.36–3.42 (m, 1 H), 3.70–3.77 (m, 2 H), 7.30–7.34 (m, 3 H), 7.51–7.55 (m, 2 H); ¹³C NMR (75.5 MHz, CDCl₃) δ –1.7 (CH₃), 5.1 (CH₂), 6.9 (CH₃), 19.6 (CH₂), 22.9 (CH₂), 33.2 (CH₂), 52.8 (CH), 60.0 (CH), 65.2 (CH), 65.3 (CH), 72.9 (CH), 82.5 (CH), 118.2 (CN), 127.6 (CH), 128.7 (CH), 133.6 (CH), 140.0 (C). Minor isomer: [α]_D²⁴ +50 (c 0.5, CHCl₃); IR (film) 2960, 2910, 2876, 2234 (w), 1957, 1453, 1246, 1148, 1007 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.38 (s, 3 H), 0.42 (s, 3 H), 0.48–0.62 (m, 12 H), 0.86–0.99 (m, 20 H), 1.65–1.74 (m, 1 H), 1.88–1.98 (m, 1 H), 2.11–2.22 (m, 2 H), 2.94 (ddd, *J* = 7.0, 7.0, 3.3 Hz, 1 H), 3.53 (ddd, *J* = 8.0, 6.4, 4.5 Hz, 1 H), 3.62 (dd, *J* = 3.3, 3.3 Hz, 1 H), 3.82 (dd, *J* = 6.8, 6.8 Hz, 1 H), 4.11 (dd, *J* = 6.4, 3.3 Hz, 1 H), 7.33–7.37 (m, 3 H), 7.52–7.57 (m, 2 H); ¹³C NMR (75.5 MHz, CDCl₃) δ –2.6 (CH₃), –1.6 (CH₃), 4.8 (CH₂), 4.9 (CH₂), 6.8 (CH₃), 21.8 (CH₂), 25.6 (CH₂), 31.8 (CH₂), 56.8 (CH), 65.3 (CH), 67.4 (CH), 73.9 (CH), 80.9 (CH), 121.4 (CN), 127.8 (CH), 128.9 (CH), 133.6 (CH), 139.2 (C); MS (ESI) *m/z* 545 (MH⁺); HRMS (ESI) calcd for C₂₉H₅₃N₂O₂Si₃ 545.3409, found 545.3413 (MH⁺). Anal. Calcd for C₂₉H₅₂N₂O₂Si₃: C, 63.91; H, 9.62; N, 5.14. Found: C, 63.81; H, 9.54; N, 4.78.

(3aR,4S,8aR,8bS)-4-((Dimethyl(phenyl)silyl)methyl)-hexahydro-[1,3]dioxolo[4,5-a]pyrrolizin-6-one (10). To a stirred solution of diol **7** (14.3 mg, 0.047 mmol) in DCM (0.22 mL) at 20 °C were added dimethoxyethane (0.22 mL) and phosphoric anhydride (31.0 mg, 0.218 mmol). After being stirred for 16 h, the reaction mixture was quenched with a cold saturated solution of sodium bicarbonate and extracted with DCM. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. Purification of the resultant crude material by flash chromatography on silica gel with ether in dichloromethane (3–

10%) provided 13.1 mg (88%) of protected diol **10**: [α]_D²⁴ –59 (c 1.0, CHCl₃); IR (film) 3068, 2954, 1690, 1407 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.36 (s, 3 H), 0.40 (s, 3 H), 0.95 (A of ABX, *J* = 14.8, 10.3 Hz, 1H), 1.02 (B of ABX, *J* = 14.8, 6.6 Hz, 1H), 1.82 (dddd, *J* = 6.2, 8.9, 9.1, 13.2 Hz, 1 H), 2.11 (dddd, *J* = 4.2, 6.7, 10.8, 13.2 Hz, 1 H), 2.26–2.39 (m, 2 H), 3.68 (ddd, *J* = 4.1, 4.1, 8.9, 1 H), 4.23 (ddd, *J* = 0.0, 6.6, 10.3 Hz, 1 H), 4.31–4.36 (m, 2 H), 4.69 (s, 1 H), 4.92 (s, 1 H), 7.33–7.37 (m, 3 H), 7.48–7.53 (m, 2 H); ¹³C NMR (125 MHz, CDCl₃) δ –3.6 (CH₃), –2.8 (CH₃), 17.1 (CH₂), 19.2 (CH₂), 32.8 (CH₂), 56.3 (CH), 61.6 (CH), 80.0 (CH), 89.6 (CH), 96.1 (CH₂), 127.7 (CH), 129.0 (CH), 133.4 (CH), 138.7 (C), 175.6 (C); MS (ESI) *m/z* 318 (MH⁺), 340 (MNa⁺), 635 (2MH⁺); HRMS (ESI) calcd for C₁₇H₂₄N₂O₃Si 318.1525, found 318.1522 (MH⁺).

(3aR,4S,6R,8aR,8bS)- and (3aR,4S,6S,8aR,8bS)-4-((Dimethyl(phenyl)silyl)methyl)-hexahydro-4H-[1,3]dioxolo[4,5-a]pyrrolizine-6-carbonitriles (11). A stirred solution of protected diol **10** (0.037 g, 0.117 mmol) in anhydrous THF (3.0 mL) was treated at 0 °C with a freshly prepared solution of DIBALH-BuLi ate complex (0.38 mL) and then TMSCN, as described for compound **9**, to give 0.045 g of crude oil (1.5:1 diastereomeric mixture), which was heated in DMF at 145 °C for 45 min. The 9:1 diastereomeric mixture obtained after removal of solvent was purified by flash chromatography on silica gel with ethyl acetate in pentane (5–15%) to give 0.033 g (86%) of the major isomer of amino-nitriles **11** and 0.0026 g (7%) of the minor isomer. Major isomer: [α]_D²⁴ +14 (c 0.25, CHCl₃); IR (film) 3063, 2953, 2922, 2225, 1096 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.35 (s, 6 H), 0.84 (A of ABX, *J* = 14.8, 9.9 Hz, 1 H), 0.86 (B of ABX, *J* = 14.8, 6.6 Hz, 1 H), 1.85–2.08 (m, 3 H), 2.12–2.25 (m, 1 H), 3.40–3.50 (m, 2 H), 3.93 (dd, *J* = 5.8, 7.9 Hz, 1 H), 4.22 (dd, *J* = 0.0, 5.6 Hz, 1 H), 4.33 (dd, *J* = 5.6, 5.6 Hz, 1H), 4.55 (s, 1 H), 4.98 (s, 1 H), 7.25–7.31 (m, 3 H), 7.44–7.52 (m, 2 H); ¹³C NMR (75.5 MHz, CDCl₃) δ –3.0 (CH₃), –2.6 (CH₃), 21.4 (CH₂), 23.9 (CH₂), 31.9 (CH₂), 54.2 (CH), 63.7 (CH), 65.7 (CH), 82.7 (CH), 90.4 (CH), 96.1 (CH₂), 121.2 (C), 127.8 (CH), 128.9 (CH), 133.6 (CH), 139.1 (C); MS (ESI) *m/z* 329 (MH⁺); HRMS (ESI) calcd for C₁₈H₂₅N₂O₂Si 329.1685, found 329.1683 (MH⁺). Minor isomer: [α]_D²⁴ –12 (c 0.32, CHCl₃); IR (film) 3073, 2953, 2926, 2238, 1090 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.33 (s, 3 H), 0.34 (s, 3 H), 1.07 (d, *J* = 7.8 Hz, 2 H), 1.79–1.89 (m, 1 H), 2.00–2.12 (m, 1 H), 2.18–2.31 (m, 2 H), 3.51 (ddd, *J* = 2.4, 7.8, 7.8 Hz, 1 H), 3.57–3.67 (m, 2 H), 4.28 (dd, *J* = 5.0, 5.0 Hz, 1H), 4.44 (dd, *J* = 2.4, 5.0 Hz, 1 H), 4.83 (s, 1 H), 5.32 (s, 1 H), 7.32–7.37 (m, 3 H), 7.48–7.53 (m, 2 H); ¹³C NMR (75.5 MHz, CDCl₃) δ –2.4 (CH₃), –2.3 (CH₃), 22.3 (CH₂), 23.5 (CH₂), 33.6 (CH₂), 52.4 (CH), 61.9 (CH), 65.7 (CH), 81.6 (CH), 91.8 (CH), 96.4 (CH₂), 119.6 (C), 127.8 (CH), 128.9 (CH), 133.4 (CH), 139.1 (C); MS (ESI) *m/z* 329 (MH⁺); HRMS (ESI) calcd for C₁₈H₂₅N₂O₂Si 329.1685, found 329.1682 (MH⁺).

(2S,3aR,4S,8aR,8bS)-4-((Dimethyl(phenyl)silyl)methyl)-2-phenylhexahydro-[1,3]dioxolo[4,5-a]pyrrolizin-6-one (12). To a stirred solution of diol **7** (0.105 g, 0.34 mmol) in DCM (3.4 mL) at 20 °C were added PhCH(OMe)₂ (0.072 mL, 0.48 mmol) and *p*-TSA-H₂O (6.4 mg, 0.03 mmol). After being stirred for 16 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ and extracted with ethyl acetate. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure to give 0.136 g of crude **12**, which was used without further purification. An analytical sample was obtained by purification of comparable crude material by flash chromatography on silica gel with ethyl acetate in pentane (1:10), which afforded **12** as a white solid: mp 134–135 °C; [α]_D²³ –72 (c 2.1, CHCl₃); IR (film) 3076, 2946, 1688, 1465 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.41 (s, 3 H), 0.46 (s, 3 H), 0.98 (B of ABX, *J* = 14.7, 10.7 Hz, 1 H), 1.06 (A of ABX, *J* = 14.7, 6.5 Hz, 1 H), 1.72–1.85 (m, 1 H), 2.10 (dddd, *J* = 13.2, 8.0, 8.0, 3.4 Hz, 1 H), 2.26–2.41 (m, 2 H), 3.72 (ddd, *J* = 8.9, 3.4, 3.4 Hz, 1 H), 4.47 (dd, *J* = 10.7, 6.5 Hz, 1 H), 4.50–4.56 (m, 2 H), 5.61 (s, 1 H), 7.31–7.45 (m, 8 H), 7.49–7.63 (m, 2 H); ¹³C NMR (100 MHz, CDCl₃) δ –3.6 (CH₃), –2.8 (CH₃), 16.7 (CH₂), 18.8 (CH₂), 32.7 (CH₂), 57.0 (CH), 61.6 (CH), 81.1 (CH), 90.1 (CH), 105.4 (CH), 127.0 (CH), 127.7 (CH), 128.5 (CH), 129.0 (CH), 129.8 (CH), 133.4 (CH), 135.4 (C), 138.8 (C), 176.4 (C); MS (ESI) *m/z* 416

(MNa⁺), 809 (2M + Na)⁺; HRMS (ESI) calcd for C₂₃H₂₇NO₃NaSi 416.1652, found 416.1646 (MNa⁺).

(2S,3aR,4S,6R,8aR,8bS)- and (2S,3aR,4S,6S,8aR,8bS)-4-((dimethyl(phenyl)silyl)methyl)-2-phenylhexahydro-4H-[1,3]-dioxolo[4,5-a]pyrrolizine-6-carbonitriles (13). A stirred solution of the above crude lactam **12** in anhydrous THF (3.0 mL) was treated at 0 °C with 1.6 mL of a freshly prepared solution of DIBALH-BuLi ate complex (prepared from 5.5 mL of 1.0 M DIBAL-H in hexanes and 2.0 mL of 2.5 M *n*-BuLi in hexanes in 2.5 mL of anhydrous THF at 0 °C). The reaction was allowed to warm to 20 °C and was monitored by TLC. After 1.5 h, the reaction mixture was cooled to 0 °C and TMSCN (0.236 mL, 0.187 g, 1.89 mmol) was added. After an additional 20 min, EtOAc and water were added and the resulting mixture was stirred for another 20 min. The mixture was then directly filtered through a plug of Celite, and the latter was washed with EtOAc. The filtrate was washed with brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The crude product was purified by flash chromatography on silica gel with ethyl acetate in pentane (2:8) to afford 0.135 g (97% overall from **7**) of a 6.7:1 mixture of amino-nitriles **13** as a colorless oil. Pure, analytical samples of each epimers could be secured by a second chromatography on silica gel. Major isomer: [α]_D²¹ +57 (c 2.2, CHCl₃); IR (film) 3063, 2919, 2237 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.44 (s, 6 H), 0.99 (d, J = 8.2 Hz, 2 H), 1.92–2.08 (m, 2 H), 2.13–2.32 (m, 2 H), 3.56–3.63 (m, 1 H), 3.65 (t, J = 8.2 Hz, 1 H), 4.08 (dd, J = 8.1, 6.3 Hz, 1 H), 4.53 (dd, J = 6.0, 0.8 Hz, 1 H), 4.61 (dd, J = 6.0, 4.9 Hz, 1 H), 5.68 (s, 1 H), 7.34–7.38 (m, 3 H), 7.54–7.60 (m, 2 H), 7.39 (s, 5 H); ¹³C NMR (100 MHz, CDCl₃) δ -2.9 (CH₃), -2.6 (CH₃), 21.4 (CH₂), 23.4 (CH₂), 31.8 (CH₂), 54.0 (CH), 63.9 (CH), 65.7 (CH), 83.7 (CH), 91.2 (CH), 104.8 (CH), 121.2 (C), 126.0 (CH), 127.7 (CH), 128.5 (CH), 128.9 (CH), 129.3 (CH), 133.6 (CH), 135.9 (C), 139.2 (C); MS (ESI) *m/z* 405 (MH⁺); HRMS (ESI) calcd for C₂₄H₂₉N₂O₂Si 405.1993, found 405.2000 (MH⁺). Minor isomer: [α]_D²¹ +15 (c 0.7, CHCl₃); IR (film) 2923, 2236, 1459, 1426, 1407, 1248, 1112 cm⁻¹; ¹H NMR (400 MHz, toluene-*d*₈) δ 0.31 (s, 6 H), 0.90 (b of ABX, J = 14.7, 7.4 Hz, 1 H), 0.97 (A of ABX, J = 14.7, 7.8 Hz, 1 H), 1.28 (dddd, J = 12.8, 6.7, 6.7, 5.6 Hz, 1 H), 1.46 (dddd, J = 12.1, 7.9, 6.7, 6.7 Hz, 1 H), 1.86 (dddd, J = 12.1, 6.7, 5.6, 5.6 Hz, 1 H), 2.01 (dddd, J = 12.8, 7.9, 6.7, 6.7 Hz, 1 H), 3.16 (ddd, J = 6.7, 6.7, 5.3 Hz, 1 H), 3.28 (dd, J = 6.7, 5.6 Hz, 1 H), 3.85 (ddd, J = 7.8, 7.4, 3.1 Hz, 1 H), 4.02 (dd, J = 5.6, 5.3 Hz, 1 H), 4.16 (dd, J = 5.6, 3.1 Hz, 1 H), 5.60 (s, 1 H), 7.17–7.25 (m, 6 H), 7.69–7.73 (m, 2 H), 7.40–7.46 (m, 2 H); ¹³C NMR (100 MHz, toluene-*d*₈) δ -2.5 (CH₃), -2.2 (CH₃), 22.4 (CH₂), 24.8 (CH₂), 33.3 (CH₂), 52.2 (CH), 63.4 (CH), 65.8 (CH), 82.1 (CH), 91.2 (CH), 106.6 (CH), 118.5 (C), 127.5 (CH), 128.0 (CH), 128.4 (CH), 129.1 (CH), 129.3 (CH), 133.8 (CH), 136.8 (C), 139.7 (C); MS (ESI) *m/z* 405 (MH⁺), 427 (MNa⁺).

(1S,2R,3S,5R,7aR)-3,5-Bis((dimethyl(phenyl)silyl)methyl)-1,2-bis(triethylsilyloxy)-hexahydro-1H-pyrrolizine (14). To the diastereomeric mixture of amino-nitriles **9** (0.201 g, 0.369 mmol) in anhydrous ether (7.0 mL) was added at 0 °C dimethylphenylsilylmethylmagnesium chloride (3.7 mL, 0.8 M in THF, 3.0 mmol). The reaction mixture was allowed to warm to 20 °C over 45 min and stirred for 48 h, after which it was cooled to 0 °C and treated with a saturated aqueous solution of NH₄Cl. The mixture was extracted with ethyl acetate, and the combined organic layers were washed with water, dried over anhydrous Na₂SO₄, and filtered. Concentration of the filtrate under reduced pressure gave the crude product **14** (5:1 diastereomeric mixture), which was used directly below. A comparable sample was subjected to silica gel chromatography for characterization purposes. Mixture of isomers: IR (film) 3068, 3048, 2953, 2876, 1426, 1249, 1112 cm⁻¹; MS (ESI) *m/z* 668 (MH⁺); HRMS (ESI) calcd for C₃₇H₆₆NO₂Si₄ 668.4165, found 668.4166 (MH⁺). Major isomer: ¹H NMR (400 MHz, CDCl₃) δ 0.21 (s, 6 H), 0.29 (s, 3 H), 0.34 (s, 3 H), 0.42–0.50 (m, 6 H), 0.52–0.59 (m, 6 H), 0.73 (dd, J = 13.7, 12.7 Hz, 1 H), 0.83–0.89 (m, 9 H), 0.90–0.95 (m, 9 H), 0.98–1.12 (m, 3 H), 1.14–1.27 (m, 1 H), 1.41–1.50 (m, 1 H), 1.66–1.75 (m, 1 H), 2.00–2.11 (m, 1 H), 2.82–2.88 (m, 1 H), 3.15–3.25 (m, 1 H), 3.50 (ddd, J = 15.3, 7.2, 7.2 Hz, 1 H), 3.66 (dd, J = 3.3, 3.3 Hz, 1 H), 4.12 (dd, J = 7.2, 3.3 Hz, 1 H), 7.29–7.35 (m, 6 H), 7.42–7.47 (m, 2 H), 7.48–7.53

(m, 2 H); ¹³C NMR (75.5 MHz, CDCl₃) δ -2.2 (CH₃), -2.1 (CH₃), -1.4 (CH₃), 4.9 (CH₂), 5.0 (CH₂), 6.8 (CH₃), 6.9 (CH₃), 23.3 (CH₂), 23.6 (CH₂), 26.4 (CH₂), 34.5 (CH₂), 65.0 (CH), 65.2 (CH), 65.8 (CH), 74.5 (CH), 80.8 (CH), 127.6 (CH), 127.7 (CH), 128.6 (CH), 128.8 (CH), 133.5 (CH), 133.7 (CH), 139.5 (C), 139.8 (C). Minor isomer (from resilylation of **15b**): ¹H NMR (400 MHz, CDCl₃) δ 0.21 (s, 3 H), 0.23 (s, 3 H), 0.28 (s, 3 H), 0.32 (s, 3 H), 0.49–0.67 (m, 12 H), 0.77 (dd, J = 14.3, 11.8 Hz, 1 H), 0.88–0.99 (m, 18 H), 1.00–1.12 (m, 3 H), 1.27–1.37 (m, 2 H), 1.45–1.54 (m, 1 H), 1.89–1.98 (m, 1 H), 2.95–3.03 (m, 1 H), 3.18 (ddd, J = 7.5, 4.1, 4.1 Hz, 1 H), 3.35 (ddd, J = 8.5, 5.3, 5.3 Hz, 1 H), 3.63 (dd, J = 4.1, 4.1 Hz, 1 H), 3.99 (dd, J = 5.3, 4.1 Hz, 1 H), 7.28–7.37 (m, 6 H), 7.43–7.47 (m, 2 H), 7.48–7.53 (m, 2 H); ¹³C NMR (75.5 MHz, CDCl₃) δ -2.3 (CH₃), -2.1 (CH₃), -1.4 (CH₃), 4.9 (CH₂), 5.2 (CH₂), 6.9 (CH₃), 7.0 (CH₃), 16.7 (CH₂), 24.1 (CH₂), 24.7 (CH₂), 32.1 (CH₂), 58.6 (CH), 59.0 (CH), 66.2 (CH), 73.3 (CH), 80.3 (CH), 127.6 (CH), 128.0 (CH), 128.6 (CH), 128.7 (CH), 133.0 (CH), 133.5 (CH), 133.7 (CH), 139.5 (C), 139.9 (C).

(1S,2R,3S,5R,7aR)- and (1S,2R,3S,5S,7aR)-3,5-Bis((dimethyl(phenyl)silyl)methyl)-hexahydro-1H-pyrrolizine-1,2-diols (15a,b). A solution of the above crude diastereomeric mixture **14** in THF at 20 °C was treated with TBAF (1.1 mL, 1.0 M in THF) and then stirred for 4 h, after which water was added and the mixture was extracted with chloroform. The combined organic layers were washed with water, dried over anhydrous Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure to give the crude diols, which were purified by flash silica gel chromatography with methanol–aqueous NH₄OH–chloroform (0.4:0.4:10) to afford 0.111 g (68% from **9**) of the major isomer **15a** and 0.019 g (12% from **9**) of the minor isomer **15b**. Minor isomer: [α]_D²⁴ -9 (c 1.1, CHCl₃); IR (film) 3378, 3072, 3048, 2953, 2910, 1113 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.24 (s, 3 H), 0.26 (s, 3 H), 0.32 (s, 3 H), 0.34 (s, 3 H), 0.73 (dd, J = 14.0, 13.0 Hz, 1 H), 1.00–1.19 (m, 3 H), 1.32–1.52 (m, 2 H), 1.64–1.71 (m, 1 H), 1.73–1.87 (m, 1 H), 2.10–2.70 (br s, 2 H), 3.06–3.26 (m, 2 H), 3.38–3.49 (m, 1 H), 3.71 (dd, J = 6.6, 4.5 Hz, 1 H), 3.85 (dd, J = 4.5, 4.5 Hz, 1 H), 7.29–7.38 (m, 6 H), 7.42–7.49 (m, 2 H), 7.50–7.57 (m, 2 H); ¹³C NMR (75.5 MHz, CDCl₃) δ -2.2 (CH₃), -1.6 (CH₃), 17.3 (CH₂), 22.2 (CH₂), 23.1 (CH₂), 32.8 (CH₂), 57.2 (CH), 57.8 (CH), 65.6 (CH), 71.3 (CH), 80.6 (CH), 127.79 (CH), 127.88 (CH), 128.9 (CH), 133.4 (CH), 133.6 (CH), 139.7 (C). Major isomer: [α]_D²⁴ +24 (c 0.9, CHCl₃); IR (film) 3388, 3068, 3048, 2952, 2908, 1426, 1248, 1113 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.28 (s, 6 H), 0.33 (s, 3 H), 0.38 (s, 3 H), 0.89 (dd, J = 14.2, 11.5 Hz, 1 H), 1.14 (dd, J = 14.7, 11.0 Hz, 1 H), 1.20–1.40 (m, 3 H), 1.61–1.71 (m, 1 H), 1.79–1.98 (m, 2 H), 2.21 (br s, 2 H), 2.71 (ddd, J = 11.0, 8.1, 3.0 Hz, 1 H), 2.84–2.91 (m, 1 H), 3.55 (ddd, J = 7.5, 7.5, 4.2 Hz, 1 H), 3.68–3.73 (m, 1 H), 3.76 (dd, J = 4.2, 4.2 Hz, 1 H), 7.32–7.39 (m, 6 H), 7.47–7.52 (m, 2 H), 7.53–7.58 (m, 2 H); ¹³C NMR (75.5 MHz, CDCl₃) δ -2.4 (CH₃), -2.0 (CH₃), -1.2 (CH₃), 23.1 (CH₂), 23.6 (CH₂), 24.0 (CH₂), 34.8 (CH₂), 64.6 (CH), 64.8 (CH), 65.5 (CH), 72.5 (CH), 80.9 (CH), 127.7 (CH), 128.0 (CH), 128.8 (CH), 129.0 (CH), 133.5 (CH), 139.4 (C), 139.7 (C); MS (ESI) *m/z* 440 (MH⁺); HRMS (ESI) calcd for C₂₅H₃₈NO₂Si₂ 440.2436, found 440.2435 (MH⁺).

(1S,2R,3S,5R,7aR)-3,5-Bis((dimethyl(phenyl)silyl)methyl)-hexahydro-1H-pyrrolizine-1,2-diol (15a from 13). To the diastereomeric mixture of amino-nitriles **13** (10.0 mg, 0.025 mmol) in anhydrous ether (0.5 mL) at 0 °C was added a solution of dimethylphenylsilylmethylmagnesium chloride (0.247 mL, 0.8 M in THF, 0.20 mmol). The reaction mixture was allowed to warm to 20 °C over 45 min and stirred for 48 h, after which it was cooled to 0 °C and treated with a saturated aqueous solution of NH₄Cl. The mixture was extracted with ethyl acetate, and the combined organic layers were washed with water, dried over anhydrous Na₂SO₄, and filtered. Concentration of the filtrate under reduced pressure gave the crude product **16**, which was used directly below. A comparable sample was subjected to silica gel chromatography for characterization purposes: IR (film) 3068, 3020, 2951, 2920, 1664, 1589, 1561, 1426, 1248, 1113, 1087 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 0.10 (s, 3 H), 0.15 (s, 3 H), 0.34 (s, 3 H), 0.36 (s, 3 H), 0.64 (dd, J = 11.6, 14.2 Hz, 1 H),

0.94–1.08 (m, 3 H), 1.19–1.32 (m, 1 H), 1.78–1.89 (m, 2 H), 2.02–2.13 (m, 1 H), 3.23–3.33 (m, 1 H), 3.37 (dd, $J = 6.5, 9.0$ Hz, 1 H), 3.66 (ddd, $J = 4.1, 4.1, 8.6$ Hz, 1 H), 4.49 (dd, $J = 0.6, 6.1$ Hz, 1 H), 4.59 (dd, $J = 5.3, 11.1$ Hz, 1 H), 5.67 (s, 1 H), 7.28–7.41 (m, 13 H), 7.50–7.55 (m, 2 H); ^{13}C NMR (75.5 MHz, CDCl_3) δ -2.4 (CH_3), -2.3 (CH_3), -2.2 (CH_3), -2.1 (CH_3), 22.3 (CH_2), 22.5 (CH_2), 23.6 (CH_3), 34.6 (CH_2), 60.9 (CH), 61.7 (CH), 64.7 (CH), 84.8 (CH), 91.1 (CH), 105.4 (CH), 126.7 (CH), 127.6 (CH), 127.7 (CH), 128.2 (CH), 128.7 (CH), 129.1 (CH), 133.4 (CH), 133.6 (CH), 136.4 (C), 139.0 (C), 139.6 (C); MS (ESI) m/z 528 (MH^+); HRMS (ESI) calcd for $\text{C}_{32}\text{H}_{42}\text{O}_2\text{NSi}_2$ 528.2749, found 528.2741 (MH^+). The above crude mixture was dissolved in 0.600 mL of TFA/ H_2O /THF (1:2:3) and heated at 60 °C for 3 h. After being allowed to cool to 20 °C, the reaction mixture was quenched by the addition of 1 N NaOH and extracted with AcOEt. The combined organic layers were washed with brine, dried over anhydrous Na_2SO_4 , and filtered. The filtrate was concentrated under reduced pressure to give the crude product, which was purified by flash chromatography on silica gel with methanol–aqueous NH_4OH –chloroform to afford 9.6 mg (88%) of pure diol 15a.

(1S,2R,3R,5R,7aR)-3,5-Bis(hydroxymethyl)-hexahydro-1H-pyrrolizine-1,2-diol (Hyacinthacine B₁ (2)). A solution of diol 15a (66 mg, 0.150 mmol) in dichloromethane (2.0 mL) at 20 °C was treated with 0.182 mL (0.200 g, 1.50 mmol) of tetrafluoroboric acid dimethyl ether complex and stirred for 5 h, whereupon the solution was concentrated under reduced pressure. A solution of the resulting crude material in dimethylformamide (1.5 mL) was treated with 86 mg (1.48 mmol) of potassium fluoride and, after 30 min, cooled to 0 °C and treated with 0.288 mL (0.101 g, 35% in water, 2.96 mmol) of aqueous hydrogen peroxide. The reaction mixture was allowed to warm to 20 °C over 1 h and then heated at 40 °C for 2 h. Solid NaHSO_3 (0.308 g) was added to the reaction mixture at 0 °C (until the starch-iodide test was negative) and the mixture was then filtered through a small pad of sand, which was washed with dimethylformamide. The filtrate was concentrated under reduced pressure at 20 °C to afford the crude product, which was purified by flash silica gel chromatography with methanol–aqueous NH_4OH –chloroform (1:1:5) and then passed through a Dowex (OH^-) ion exchange column with water to afford 23 mg (75%) of pure hyacinthacine B₁ (2): $[\alpha]_{\text{D}}^{24} +40$ (c 0.4, H_2O); IR (film) 3352, 2928, 2880 cm^{-1} ; ^1H NMR (400 MHz, CD_3OD) δ 1.52–1.61 (m, 1 H), 1.73–1.82 (m, 1 H), 2.00–2.10 (m, 1 H), 2.10–2.20 (m, 1H), 2.91 (ddd, $J = 9.0, 6.1, 3.1$ Hz, 1 H), 3.03–3.10 (m, 1 H), 3.42 (dd, $J = 11.0, 6.4$ Hz, 1 H), 3.44 (dd, $J = 11.0, 5.4$ Hz, 1H), 3.54–3.58 (m, 1 H), 3.60 (dd, $J = 10.9, 6.1$ Hz, 1H), 3.78 (dd, $J = 10.9, 3.1$ Hz, 1 H), 3.83 (dd, $J = 3.8, 3.8$ Hz, 1 H), 3.92 (dd, $J = 8.8, 4.1$ Hz, 1 H); ^1H NMR (400 MHz, D_2O) δ 1.46–1.55 (m, 1 H), 1.73–1.82 (m, 1 H), 1.91–2.08 (m, 2 H), 2.85–2.92 (m, 1 H), 2.97–3.05 (m, 1 H), 3.42 (dd, $J = 11.5, 6.0$ Hz, 1 H), 3.49–3.54 (m, 1 H), 3.55 (dd, $J = 11.0, 6.5$ Hz, 1 H), 3.60 (dd, $J = 11.5, 6.5$ Hz, 1 H), 3.73 (dd, $J = 11.5, 4.0$ Hz, 1 H), 3.90–3.95 (m, 2 H); ^{13}C NMR (100.6 MHz, CD_3OD) δ 25.0 (CH_2), 31.2 (CH_2), 65.6 (CH_2), 67.4 (CH_2), 69.3 (CH), 71.4 (CH), 72.4 (CH), 74.6 (CH), 77.6 (CH); ^{13}C NMR (100.6 MHz, D_2O) δ 25.3 (CH_2), 32.3 (CH_2), 66.1 (CH_2), 67.5 (CH_2), 69.4 (CH), 71.9 (CH), 72.1 (CH), 74.9 (CH), 78.2 (CH); MS (ESI) m/z 204 (MH^+); HRMS (ESI) calcd for $\text{C}_9\text{H}_{18}\text{NO}_4$ 204.1230, found 204.1230 (MH^+).

(2R,3S,4R,5R)-2-(But-3-enyl)-5-(hydroxymethyl)pyrrolidine-3,4-diol (17). A solution of diol 15b (16.0 mg, 0.036 mmol) in dichloromethane (1.0 mL) was stirred with 0.044 mL (48 mg, 0.36 mmol) of tetrafluoroboric acid dimethyl ether complex for 5 h at 20 °C. The solvent was then evaporated, and the resulting crude product was dissolved in DMF (0.5 mL) and treated with 21 mg (0.36 mmol) of potassium fluoride. After being stirred for 30 min, the reaction mixture was cooled to 0 °C, and 0.070 mL (24 mg, 35% in water, 0.72 mmol) of hydrogen peroxide solution was added. After being allowed to warm to 20 °C over 1 h, the reaction mixture was heated at 40 °C for 2 h. The excess peroxide was destroyed by addition of solid NaHSO_3 (75 mg) at 0 °C, and the resulting reaction mixture was filtered through a small pad of sand, which was then washed with DMF. The solvents were evaporated under reduced pressure at 20 °C to afford the crude

product, which was purified by flash chromatography on silica gel with methanol–aqueous NH_4OH –chloroform (1:1:5). This resulting salt, after passage over Dowex (OH^-) with water, gave 4.7 mg (69%) of pure 17: ^1H NMR (400 MHz, D_2O) δ 1.53–1.64 (m, 1 H), 1.65–1.76 (m, 1 H), 2.14 (q, $J = 7.6, 14.4$ Hz, 2 H), 3.09–3.20 (m, 2 H), 3.63 (dd, $J = 6.4, 12.0$ Hz, 1 H), 3.73 (dd, $J = 4.2, 11.3$ Hz, 1 H), 4.01 (dd, $J = 4.2, 4.2$ Hz, 2 H), 4.04 (dd, $J = 4.2, 6.2$ Hz, 1 H), 5.07 (m, 2H), 5.92 (dddd, $J = 6.6, 6.6, 10.1, 10.1$ Hz, 1 H); ^{13}C NMR (75.5 MHz, D_2O) δ 28.1 (CH_2), 30.4 (CH_2), 58.8 (CH), 61.5 (CH), 62.9 (CH_2), 73.0 (CH), 74.5 (CH), 114.9 (CH_2), 139.1 (CH); MS (ESI) m/z 188 (MH^+); HRMS (ESI) calcd for $\text{C}_9\text{H}_{18}\text{O}_3\text{N}_1$ 188.1281, found 188.1278 (MH^+).

(2S,3aR,4S,6S,8aR,8bS)-6-(Benzyloxymethyl)-4-((dimethyl(phenyl)silyl)methyl)-2-phenylhexahydro-4H-[1,3]dioxolo[4,5-a]pyrrolizine (19). To a stirred solution of $^i\text{Pr}_2\text{NH}$ (0.770 mL, 0.556 g, 5.49 mmol) in freshly distilled THF (2.2 mL) at -35 °C under Ar was added dropwise 2.0 mL of n -BuLi (2.5 M in hexanes, 5.0 mmol). The solution was allowed to warm to 0 °C over 30 min and then degassed by three freeze–thaw cycles. A 0.440-mL (0.44 mmol) portion of this LDA solution was added to a degassed solution of amino-nitriles 13 (89 mg, 0.22 mmol) in THF (1.1 mL) at -60 °C, and the mixture was stirred for 15 min. Benzyl chloromethyl ether (0.093 mL, 0.105 g, 0.67 mmol) was then added at -60 °C, and stirring was continued for 30 min before the addition of saturated aqueous NH_4Cl . The reaction mixture was allowed to warm to 20 °C and was extracted with ethyl acetate. The combined organic phases were washed with water and brine, dried over anhydrous Na_2SO_4 , filtered, and concentrated under reduced pressure. The resulting, unstable crude product 18 was directly used without purification: ^1H NMR (400 MHz, toluene- d_8) δ 0.31 (s, 3 H, H), 0.34 (s, 3 H), 0.89 (B of ABX, $J = 14.6, 9.9$ Hz, 1 H), 1.12 (A of ABX, $J = 14.6, 5.0$ Hz, 1 H), 1.36–1.45 (m, 1 H), 1.64–1.75 (m, 1 H), 2.05–2.14 (m, 1 H), 2.17–2.25 (m, 1 H), 2.98 (B of AB, $J = 9.1$ Hz, 1 H), 3.20 (A of AB, $J = 9.1$ Hz, 1 H), 3.40 (ddd, $J = 6.9, 6.9, 5.0$ Hz, 1 H), 4.03 (dd, $J = 5.9, 5.0$ Hz, 1 H), 4.05 (ddd, $J = 9.9, 5.0, 3.2$ Hz, 1 H), 4.19 (dd, $J = 5.9, 3.2, 1$ H), 4.22 (B of AB, $J = 12.2$ Hz, 1 H), 4.27 (A of AB, $J = 12.2$ Hz, 1 H), 5.60 (s, 1 H), 7.16–7.27 (m, 10 H), 7.43–7.49 (m, 3 H), 7.73–7.78 (m, 2 H); ^{13}C NMR (100 MHz, toluene- d_8) δ -2.4 (CH_3), -2.2 (CH_3), 24.1 (CH_2), 24.3 (CH_2), 37.6 (CH_2), 64.1 (CH), 66.0 (C), 66.9 (CH), 73.57 (CH_2), 76.4 (CH_2), 82.2 (CH), 90.8 (CH), 106.9 (CH), 120.0 (C), 127.8 (CH), 127.9 (CH), 128.1 (CH), 128.3 (CH), 128.5 (CH), 128.6 (CH), 129.2 (CH), 129.5 (CH), 134.0 (CH), 136.7 (C), 138.3 (C), 139.6 (C); MS (ESI) m/z 525 (MH^+). To a solution of the crude product in anhydrous MeOH (1.0 mL) at 0 °C was added NaBH_4 (42 mg, 1.11 mmol). The reaction mixture was stirred for 1 h at this temperature, and then water was added and the aqueous phase was extracted with ethyl acetate. The combined organic phases were washed with brine, dried over anhydrous Na_2SO_4 , filtered, and concentrated under reduced pressure. The residue was purified by chromatography on silica gel with MeOH saturated with NH_3 in EtOAc (2–10%) to give 54 mg (49%, 55% brsm, 2 steps) of ether 19, along with 9 mg of amino-nitrile 13. Ether 19: $[\alpha]_{\text{D}}^{25} +15$ (c 1.2, CHCl_3); IR (film) 3373, 2918, 2845, 1665, 1455, 1110 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ 0.29 (s, 3 H), 0.33 (s, 3 H), 0.87 (A of ABX, $J = 14.5, 9.4$ Hz, 1 H), 1.10 (B of ABX, $J = 14.5, 5.9$ Hz, 1 H), 1.53–1.66 (m, 1 H), 1.66–1.77 (m, 1 H), 1.82–1.98 (m, 1 H), 3.08–3.16 (m, 1 H), 3.20 (B of ABX, $J = 9.1, 6.1$ Hz, 1 H), 3.38 (A of ABX, $J = 9.1, 6.2$ Hz, 1 H), 3.33–3.46 (m, 1 H), 3.60 (dd, $J = 9.4, 5.9$ Hz, 1 H), 4.26 (B of AB, $J = 12.1$ Hz, 1 H), 4.31 (A of AB, $J = 12.1$ Hz, 1 H), 5.73 (s, 1 H), 4.43–4.50 (m, 2 H), 7.27–7.40 (m, 10 H), 7.47–7.57 (m, 5 H); ^{13}C NMR (75 MHz, CDCl_3) δ -2.4 (CH_3), -2.3 (CH_3), 17.0 (CH_2), 22.0 (CH_2), 31.3 (CH_2), 56.6 (CH), 58.1 (CH), 66.5 (CH), 73.2 (2 CH_2), 80.8 (CH), 91.8 (CH), 105.4 (CH), 126.9 (CH), 127.5 (CH), 127.7 (CH), 127.9 (CH), 128.3 (CH), 128.4 (CH), 128.9 (CH), 129.2 (CH), 133.7 (CH), 137.2 (C), 138.7 (C), 139.7 (C); MS (ESI) m/z 500.2 (MH^+); HRMS (ESI) calcd for $\text{C}_{31}\text{H}_{38}\text{NO}_3\text{Si}$ 500.2615, found 500.2618 (MH^+).

(2S,3aR,4R,6S,8aR,8bS)-6-(Benzyloxymethyl)-2-phenylhexahydro-4H-[1,3]dioxolo[4,5-a]pyrrolizine-4-yl)methanol (20). 30% KH in mineral oil (27 mg, 0.20 mmol) under Ar was washed with

pentane and DMF (0.250 mL) was added. The mixture was cooled to 0 °C and carefully treated with a solution of ^tBuOOH (0.021 mL, 6.0 M in decane, 0.13 mmol) and, after 10 min, with a solution of **19** (10.5 mg, 0.021 mmol) in DMF (0.150 mL) and a solution of TBAF (0.042 mL, 1.0 M in THF, 0.04 mmol). After 1 h at 0 °C, the reaction mixture was allowed to warm to 20 °C and was stirred for 1 h. Saturated aqueous Na₂S₂O₃ was then added, and the aqueous phase was extracted with EtOAc. The combined organic phases were washed with water and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure to afford the crude product. Chromatography on silica gel with MeOH saturated with NH₃ in dichloromethane (1:10) afforded 11 mg (>100%) of alcohol **20**: [α]_D²⁵ +35.7 (c 0.8, CHCl₃); IR (film) 3375, 2926, 2853, 1674, 1403, 1086 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.62–1.81 (m, 2 H), 1.81–1.92 (m, 1 H), 2.01–2.16 (m, 1 H), 3.31–3.40 (m, 2 H), 3.40–3.47 (m, 1 H), 3.51–3.62 (m, 2 H), 3.66 (ddd, *J* = 6.3, 4.6, 1.5 Hz, 1 H), 3.70–3.78 (m, 1 H), 4.28 (B of AB, *J* = 11.9 Hz, 1 H), 4.34 (A of AB, *J* = 11.9 Hz, 1 H), 4.57–4.64 (m, 2 H), 5.84 (s, 1 H), 7.27–7.45 (m, 8 H), 7.47–7.55 (m, 2 H); ¹³C NMR (100 MHz, CDCl₃) δ 23.8 (CH₂), 30.6 (CH₂), 60.9 (CH), 62.4 (CH₂), 63.6 (CH), 67.2 (CH), 71.8 (CH₂), 73.2 (CH₂), 82.6 (CH), 86.8 (CH), 105.6 (CH), 126.5 (CH), 127.7 (CH), 127.8 (CH), 128.4 (CH), 128.5 (CH), 129.3 (CH), 136.6 (C), 138.2 (C); MS (ESI) *m/z* (MH⁺); HRMS (ESI) calcd for C₂₃H₂₈NO₄ 382.2013, found 382.2017 (MH⁺).

(+)-Hyacinthacine B₂ (3). To a solution of the above alcohol **20** in ethanol (0.300 mL) and THF (0.050 mL) were added 10% Pd/C (2.0 mg) and 6 N HCl (5 drops). The reaction mixture was stirred at 20 °C for 16 h under hydrogen. The mixture was then filtered through a plug of Celite, which was washed with MeOH. The filtrate was concentrated under reduced pressure, and the residue was placed on a column of Dowex resin (H⁺ form), which was washed successively with MeOH, water, and concentrated aqueous NH₄OH. The aqueous NH₄OH fractions were concentrated under reduced pressure to give 5.2 mg of protonated hyacinthacine B₂: ¹H NMR (400 MHz, D₂O) δ 1.81 (m, 2H, H₆ and H₇), 1.91 (m, 1H, H₆), 2.07 (m, 1H, H₇), 3.51–3.58 (m, 1H, H₃), 3.59–3.66 (m, 1H, H₅), 3.71 (B of ABX, *J* = 12.8, 2.4 Hz, 1H, H₈), 3.74 (s, 1H, H_{9b}), 3.76 (s, 1H, H_{9a}), 3.91 (A of ABX, *J* = 12.8, 3.1 Hz, 1H, H₈), 4.06–4.13 (m, 2H, H₁ and H₂), 4.15 (ddd, *J* = 8.5, 4.3, 4.1 Hz, 1H, H_{7a}); ¹³C NMR (100 MHz, D₂O) δ 25.0 (CH₂), 29.9 (CH₂), 58.2 (CH₂), 60.2 (CH₂), 64.6 (CH), 69.3 (CH), 71.8 (CH), 72.8 (CH), 73.6 (CH). This material was placed on a column of Dowex resin (OH⁻ form), which was then washed with water to give 3.8 mg (89%, 2 steps) of (+)-hyacinthacine B₂ (**3**): [α]_D²⁵ +25 (c 0.32, H₂O); IR (film) 3344, 2925, 2883, 1661, 1401, 1035 cm⁻¹; ¹H NMR (400 MHz, D₂O) δ 1.72–1.84 (m, 1 H), 1.94–2.09 (m, 3 H), 3.27 (ddd, *J* = 8.6, 6.4, 4.3 Hz, 1 H), 3.33 (pseudo-q, *J* = 5.1 Hz, 1 H), 3.66 (ddd, *J* = 7.0, 7.0, 3.9 Hz, 1 H), 3.73 (dd, *J* = 11.5, 6.4 Hz, 1 H), 3.81–3.91 (m, 3 H), 4.07 (dd, *J* = 3.9, 3.9 Hz, 1 H), 4.11 (dd, *J* = 8.6, 3.9 Hz, 1 H); ¹³C NMR (100 MHz, D₂O) δ 79.4 (CH), 74.6 (CH), 69.3 (CH), 66.8 (CH₂), 64.5 (CH₂), 64.3 (CH), 64.2 (CH), 33.1 (CH₂), 25.0 (CH₂); MS (ESI) *m/z* 204 (MH⁺); HRMS (ESI) calcd for C₉H₁₈NO₄ 204.1230, found 204.1229 (MH⁺).

■ ASSOCIATED CONTENT

■ Supporting Information

X-ray crystallographic analysis data for compound **12**, ¹H, ¹³C, and COSY NMR spectra of compounds **2**, **3**, **8–17**, **19**, and **20**, and NOESY spectra of compounds **9** (minor), **14**, and **15a**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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(19) The lactam was obtained as a 93:7 (¹H NMR) mixture of diastereomers. The minor component filtered out over the remainder of the synthesis.

(20) The analysis was performed on a racemic sample that was obtained by synthesis from racemic Stericol. After basic analysis, the crystal appeared to be monoclinic with a P₂₁ space group, which was unexpected since the product was racemic and P₂₁ is a non-centrosymmetric space group. A closer look revealed, however, that the crystal was a racemic twin with two domains and BASF = 0.58501. This confirmed that the product was truly racemic and explained some meaningless residual electronic density peaks. Crystal data for C₂₃H₂₇NO₃Si: Monoclinic, P₂₁, a = 12.493(3) Å, b = 6.4889(13) Å, c = 13.275(3) Å, β = 97.91(3)°, V = 1065.9(4) Å³, Z = 2, d_{calcd} = 1.226 mg/m³, F(000) = 420, λ Mo Kα = 0.71073 Å, Θ_{max} range 3.4–25°, 17796 measured reflections, 3624 [R(int) = 0.0338] independent reflections, R(1) [I > 2σ(I)] = 0.0800, wR2 [all data] = 0.0970, GOF (all data) = 1.099.

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(35) In order to study the rotation around the C5–CH₂Si bond (Figure 3), potential energy surface (PES) scans of **15a** and **15b** were performed at the semiempirical level of theory (PM6). Geometry optimizations were done by freezing the dihedral angle N–C5–CH₂–Si at different values between –180 and 180 degrees. Two minima with a weak energy difference and a quite flat PES were found for **15a**, whereas only one minimum was identified for **15b**. To support these results, new calculations at a high level of theory (B3lyp/6-311++G***) were performed. For **15a**, using the two minima obtained at the PM6 level as a starting point, geometry optimizations converged toward structures close to the starting point. The electronic energy values of the two minima differed by only 2.6 kcal/mol. A transition state between these two minima was localized. Its electronic energy was 3.1 kcal/mol higher than the more stable minimum, allowing rotation around the C5–CH₂Si bond for this isomer. For **15b**, the geometry optimization still converged to a minimum close to the structure obtained at the PM6 level (dihedral angle: –176.3 degrees). Other optimizations were performed with different dihedral angle values. The energy of these optimized structures increased gradually, suggesting that rotation is not energetically favorable (for example, a 12.1 kcal/mol energy difference was obtained between the minimum and the structure with a dihedral angle of –60 degrees).

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